

Involvement of hydrogen sulphide and sulphur-containing compounds in the pathogenesis and therapy of rheumatic disorders

Background

Sulphur bath therapy has long been in use for the therapy of patients suffering from rheumatic disorders and is still considered helpful for the treatment of diseases such as rheumatoid arthritis (RA) or osteoarthritis (OA). However, scientific investigations dealing with the beneficial as well as adverse effects of this kind of treatment are rare and have sometimes led to controversial results (1). Moreover, the underlying molecular mechanisms are poorly understood (2, 3). In vitro, H₂S exerts a host of effects on various biological targets, resulting in responses that range from cytotoxicity (4, 5) to cytoprotective effects (6). Several studies have demonstrated cytoprotective effects of H₂S at micromolar concentrations, which may be related to its ability to neutralize a variety of reactive species including oxyradicals (7), peroxyxynitrite (8), hypochlorous acid (9) and homocysteine (10). Exposure to higher concentrations (millimolar) of H₂S tends to be cytotoxic due to free radical and oxidant generation, calcium mobilization (11), glutathione depletion (5), as well as the induction of mitochondrial cell death pathways (4, 12).

Our own data obtained in the past three years suggest that H₂S may have anti- as well as pro-inflammatory properties depending on concentration and cell type (13-15). At high concentrations (0.5 – 1.0 mM NaHS), H₂S upregulated pro-inflammatory genes (e.g. IL-6, IL-8 and COX-2) in fibroblast-like synoviocytes (FLS) derived from RA and OA patients (15). In contrast, H₂S shows pronounced anti-inflammatory properties in monocytes and macrophages (manuscript in preparation).

Effects of hydrogen sulphide on the inflammatory status of fibroblast-like synoviocytes of rheumatoid arthritis and osteoarthritis patients

Kloesch B, Liszt M, Krehan D, Broell J, Kiener H, Steiner G (2012) Immunol Lett 141:197-203.

To explore the effects of H₂S, two FLS lines (one from a patient with RA and one from a patient with OA) were incubated for 20 min with 1.0 mM of the H₂S-donor NaHS. After changing the culture medium, incubation was continued for 12 h. At different time points, cell culture supernatants were collected and IL-6 release was quantified by ELISA. IL-6 expression was significantly increased by H₂S treatment. Furthermore, quantitative real-time PCR (qRT-PCR) revealed that in RA-FLS also IL-8, COX-2 and MMP-3 were also upregulated by NaHS treatment (Fig. 1, left panel), whereas MMP-2 and MMP-14 were negatively regulated (Fig. 1, right panel).

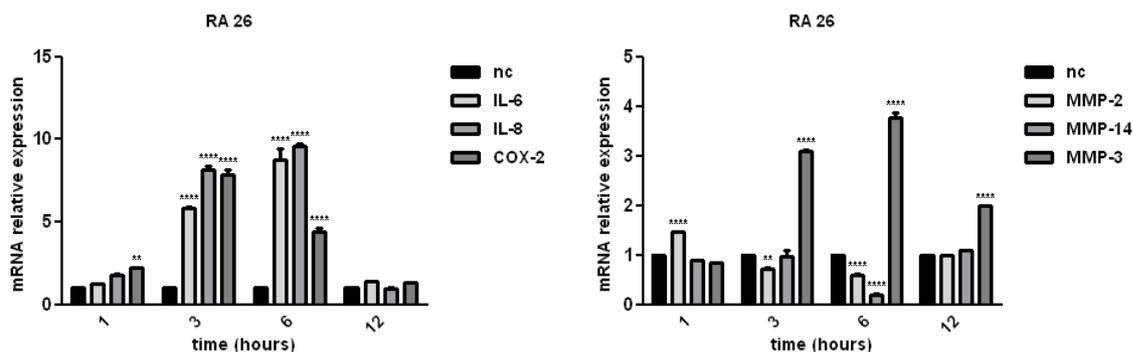


Fig. 1: H₂S upregulates mRNA levels of IL-6, IL-8, COX-2 (left panel) and MMP-3 (right panel) in RA-FLS. 1, 3, 6 and 12 h after initial H₂S exposure, total RNA was isolated and mRNA levels were quantified by qRT-PCR. Significant changes are indicated by asterisks: **p<0.01, ***p<0.0001.

To clarify the underlying mechanism leading to the induction of expression of pro-inflammatory genes by H₂S, phosphorylation of extracellular signal-regulated kinase (ERK1/2) was analyzed by Western blotting in RA and OA-FLS; already 15 min after initial H₂S exposure ERK1/2 was activated in both cell lines.

Inhibitors of p38 and ERK1/2 MAPK, SB203580 and U0126, respectively, and of NF- κ B (BAY-117082) completely blocked H₂S-induced IL-6 expression.

Taken together, these results demonstrate that high concentrations of H₂S can stimulate gene expression of IL-6 and other pro-inflammatory genes such as IL-8 and COX-2 as well as MMP-3 in FLS from RA and OA patients via activation of p38 and ERK1/2 MAPK and of NF- κ B pathway.

Effects of hydrogen sulfide on the inflammatory status of LPS-stimulated monocytes

During synovial inflammation, monocytes/macrophages play a pivotal role by synthesis and release of cytokines and chemokines important in induction and amplification of the inflammatory response. Hence, we wanted to investigate whether H₂S would prevent or rather stimulate the activation of monocytes by lipopolysaccharides (LPS) and whether NF- κ B and MAPKs were involved in this process.

The human pro-monocyte cell line U937 was differentiated for 24 h with PMA and then treated for 30 min with increasing concentrations of NaHS (0.125 – 1.0 mM) before being stimulated for 6 h with LPS (100 ng/ml). A significant reduction in IL-6 expression was obtained at 0.25 mM NaHS (Fig. 2). The highest NaHS concentration used in the experiments reduced IL-6 production at about 30 – 40 % (Fig. 1, left panel). Similar results were obtained when TNF- α release was monitored (data not shown). Notably, an almost complete suppression (~ 80 %) of IL-6 release was observed when the cells were treated 3 h after initial LPS treatment with the same dose of NaHS again (Fig. 2, right panel).

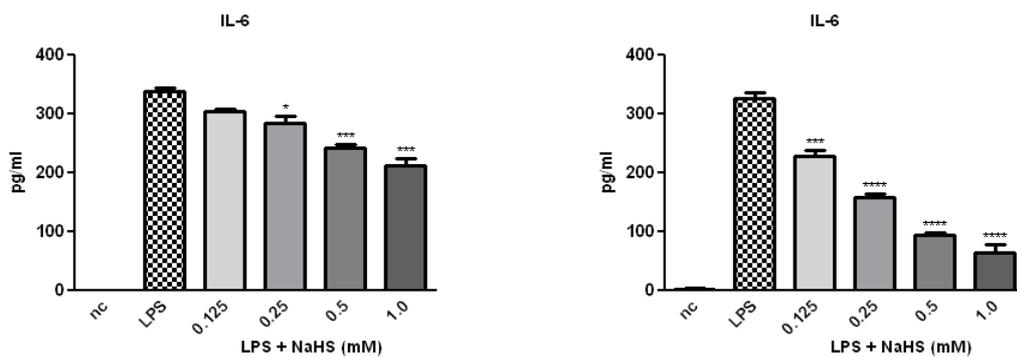


Fig. 2: H₂S blocks IL-6 expression in U937 cells. Cells remained untreated or were incubated for 6 h with LPS in the absence or presence of increasing concentrations of NaHS. IL-6 release was quantified by ELISA. Significant changes are indicated by asterisk: *p<0.05, ***p<0.001.

Remarkably, in contrast to the inhibitory effects of H₂S on TNF- α and IL-6 expression, H₂S stimulated the expression of IL-1 β (Fig. 3). This stimulation was only evident when LPS plus NaHS were present in the culture medium while NaHS alone was not able to induce IL-1 β expression (data not shown).

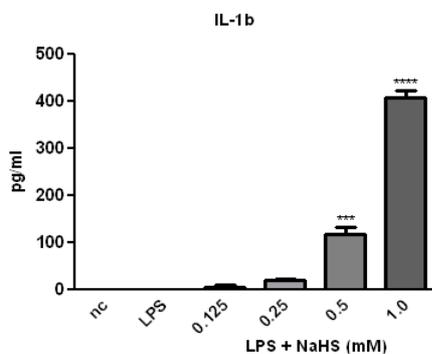


Fig. 3: H₂S induces the release of IL-1 β in U937 cells. Cells remained untreated or were incubated for 6 h with LPS in the absence or presence of increasing concentrations of NaHS. IL-1 β release was quantified by ELISA. Significant changes are indicated by asterisk: ***p<0.001, ****p<0.0001.

ersial effects in LPS-stimulated monocytes. On the one hand, H₂S ; by inhibition of TNF- α and IL-6 expression, on the other hand, it induced phosphorylation of ERK1/2 (data not shown) and stimulated the expression or the release (possibly via caspase-1 activation) of IL-1 β . Further studies are necessary to clarify the involvement of

H₂S in p38 and ERK1/2 MAPK activation/deactivation as well as the impact of H₂S on transcriptional and post-translational processes of pro-inflammatory gene expression in activated monocytes.

Effects of hydrogen sulfide and the sulphur-containing anti-oxidants dimethyl sulphoxide and dimethyl sulphone on IL-6 and IL-8 expression in a human chondrocyte cell line (C-28/I2)

Kloesch B, Liszt M, Broell J, Steiner G (2011). *Life Sci* 89:473-8.

Kloesch B, Liszt M, Steiner G, Broell J (2012) *Rheumatol Int* 32:729-36.

Next, we were interested to study the effects of H₂S on chondrocytes which are centrally involved in cartilage synthesis and metabolism. In addition, we studied the effects of two sulfur containing anti-oxidants dimethyl sulphoxide (DMSO) and dimethyl sulphone (DMS). DMSO is a powerful water miscible solvent that dissolves most water insoluble drugs. DMSO possesses anti-inflammatory properties, as well as the ability to act as a free radical scavenger (16, 17). Furthermore, it is capable of inducing or inhibiting cell proliferation, apoptosis and/or differentiation. Thus, its properties have been exploited in the treatment of dermatological, rheumatic, and renal manifestations of amyloidosis. However, limited data are available on the underlying molecular mechanism. DMS is found in small amounts in many foods, including unpasteurized milk, grains, meat, eggs, and fish. It is also present in popular dietary supplements. Reported effects claimed to be associated with DMS include relief of pain, reduction of inflammation, arthritis, allergies and asthma (18, 19).

We studied the effects of NaHS, DMSO and DMS on IL-6 and IL-8 expression in C-28/I2 cells, a human chondrocyte cell line, originally derived from a young patient with OA (20). In the case of H₂S treatment, cells were left untreated or were incubated for 15 min with different concentrations of NaHS (0.125 and 1.0 mM). After changing the culture medium, incubation was continued for 12 h. In contrast to H₂S treatment, cells were incubated in the presence of DMSO (0.5 and 1.0 %) or DMS (10 and 100 mM) over a total period of 12 h. At different time points, cell culture supernatants were collected and IL-6 and IL-8 levels were quantified by ELISAs. Reduced levels of IL-6 and IL-8 were detected from 1 to 6 h after initial H₂S exposure (14). DMSO (1.0 %) and DMS (100 mM) were much more efficient in blocking IL-6 expression than H₂S: after 12 h of incubation almost 70 % inhibition was obtained with both substances.

Taken together, these studies show that NaHS and the sulphur containing anti-oxidants DMSO and DMS are potent inhibitors of constitutive as well as IL-1 β -induced cytokine expression in a human chondrocyte cell line (21). Identification of the components of the signal transduction pathways that are sensitive to anti-oxidants may eventually open a new territory to more selective treatment of inflammatory disorders.

Effects of the polyphenols curcumin and resveratrol on cytokine expression in fibroblast-like synoviocytes

Curcumin (diferuloylmethane) is a yellow pigment found in the rhizome of turmeric (*Curcuma longa* L. *Zingiberaceae*) which has a wide range of pharmacological and biological activities (22). The anti-oxidant, anti-carcinogenic, anti-inflammatory and apoptotic effects of this compound have been assessed in various *in vitro* and *in vivo* systems. Although several studies have reported that curcumin may modulate numerous aspects of cell function relevant to inflammatory arthritis, the underlying molecular mechanisms are incompletely understood (23).

Resveratrol (*trans*-3,4,5-trihydroxystilbene) is a natural phytoalexin found in large quantities in grapes and other food products. Resveratrol was found to have a potent anti-carcinogenic activity in several animal models of cancer (24). The anti-carcinogenic properties of resveratrol are closely associated with its antioxidant activity and ability to inhibit cyclo-oxygenase, hydroperoxidase, protein kinase C, Bcl-2 phosphorylation, Akt, NF- κ B, MMP -9, and programmed cell death (25, 26).

Since little is known about the effects of curcumin and resveratrol on FLS derived from RA patients, we investigated possible anti-inflammatory and apoptotic properties of these substances in FLS obtained from two RA patients. Particular attention was paid to the influence of curcumin and resveratrol on IL-6 and VEGF-A expression in IL-1 β and PMA-stimulated FLS, as well as to its modulation of NF- κ B and MAPK such as p38 and ERK1/2.

FLS were stimulated with IL-1 β (10 ng/ml) or treated with IL-1 β plus increasing concentrations of curcumin (12.5 – 50 μ M) or resveratrol (12.5 – 100 μ M). As shown in Fig. 4, even low concentrations of curcumin (12.5 μ M) significantly blocked IL-6 release in FLS. Higher concentrations (25 – 50 μ M) almost completely suppressed IL-6 expression.

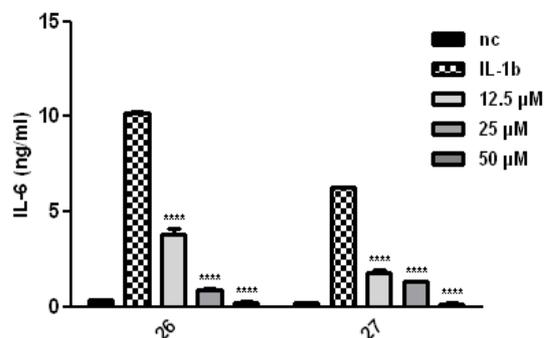


Fig. 4: Curcumin blocks IL-1 β -induced IL-6 expression in FLS. Cells remained untreated or were stimulated with IL-1 β or with IL-1 β plus increasing concentrations of curcumin. IL-6 release was quantified by ELISA (nc, negative control; 26, RA-26; 27, RA-27). Significant changes are indicated by asteriks: *** p <0.001, **** p <0.0001.

Resveratrol had much lower inhibitory capacity on IL-1 β -induced IL-6 expression compared to curcumin, and in fact, a concentration of 100 μ M resveratrol was required to obtain the same degree of suppression of IL-6 production as observed with 12.5 μ M curcumin (Fig. 5).

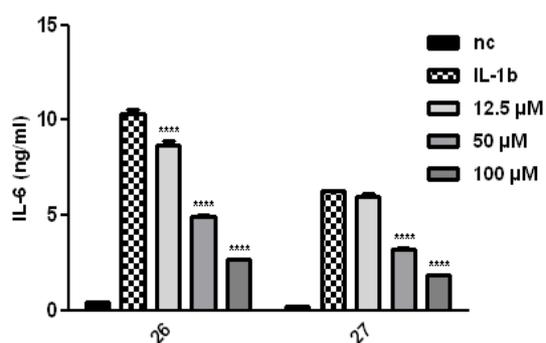


Fig. 5: Resveratrol blocks IL-1 β -induced IL-6 expression in FLS. Cells remained untreated or were stimulated with IL-1 β or with IL-1 β plus increasing concentrations of resveratrol. IL-6 release was quantified by ELISA (nc, negative control; 26, RA-26; 27, RA-27). Significant changes are indicated by asteriks: **** p <0.0001.

Phorbol esters such as PMA stimulate protein kinase C and activate the Ras/Raf/ERK1/2 signalling pathway. Therefore the capacity of curcumin and resveratrol to block PMA-induced IL-6 expression was studied. The results obtained were similar to those obtained with IL-1 β stimulated cells (data not shown).

To investigate the influence of curcumin and resveratrol on MAPK activation, FLS were incubated for 30 min with increasing concentrations of curcumin (12.5 – 50 μ M) or resveratrol (12.5 – 100 μ M) before being stimulated for 20 min with IL-1 β . As shown in Fig. 6, both curcumin and resveratrol induced dephosphorylation of ERK1/2 in a concentration-dependent manner; however, p38 MAPK activation was unaffected by either substance.

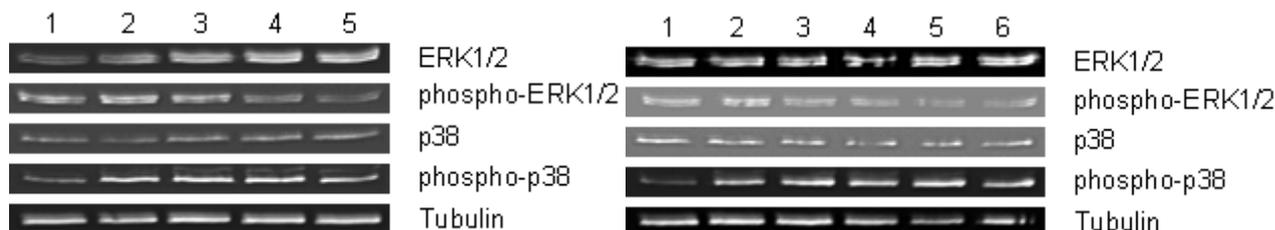


Fig. 6: Curcumin and resveratrol induce ERK1/2 deactivation in FLS. Cells remained untreated or were incubated for 30 min with increasing concentrations of curcumin (left panel) or resveratrol (right panel) before being stimulated for 20 min with IL-1 β (1, negative control; 2, IL-1 β ; 3, 12.5 μ M curcumin/resveratrol; 4, 25 μ M curcumin/resveratrol; 5, 50 μ M curcumin/resveratrol; 6, 100 μ M resveratrol).

Taken together, curcumin and resveratrol effectively suppressed IL-1 β and PMA-induced IL-6 and VEGF-A expression in FLS. Higher concentrations of curcumin (50 μ M) induced activation of caspases which finally led to induction of apoptosis whereas resveratrol did not (data not shown). Although further studies are needed, these results provide important new insights into the biological effects of curcumin and resveratrol, and suggest both substances as natural remedies for the treatment of chronic inflammatory diseases like RA.

Summary

Rheumatoid arthritis (RA) is a systemic autoimmune disease characterized by chronic inflammation of the synovial membrane, hyperplasia of the synovial lining and overactivation of osteoclasts, resulting in the irreversible destruction of articular cartilage and bone. Recently, several studies have highlighted the importance of H₂S in inflammation. H₂S suppresses leukocyte adherence to the vascular endothelium and migration of leukocytes into the subendothelial space, as well as reducing plasma exudation. H₂S has been shown to reduce expression of many pro-inflammatory cytokines, chemokines, and enzymes, most likely related to its ability to suppress activation of nuclear transcription factor- κ B (NF- κ B). H₂S is also a potent anti-oxidant and can induce apoptosis in neutrophils.

Fibroblast-like synoviocytes (FLS), located in the intimal lining layer of the synovial membrane, have been shown to promote secondary synovitis by the release of a series of pro-inflammatory cytokines (IL-6, IL-8, TNF- α) as well as matrix-metalloproteinases (MMPs).

It was our aim to investigate possible anti-inflammatory effects of exogenous H₂S (NaHS) on FLS derived from patients with RA and OA. We showed that low concentrations of H₂S (μ M) may have anti-inflammatory properties whereas high concentrations of H₂S (mM) can stimulate expression of IL-6 and other pro-inflammatory genes such as IL-8 and COX-2 as well as MMP-3. Therefore, we conclude that H₂S is a gaseous transmitter molecule with bivalent properties. Evaluation of H₂S-releasing drugs in an *in vivo* setting, i.e the collagen-induced arthritis model, will provide insight as to whether or not the exploitation of H₂S as a therapeutic agent will live up to the promise.

Beside H₂S, naturally occurring anti-oxidants such as DMS, and the polyphenols curcumin and resveratrol get an increasing importance in the treatment of chronic inflammatory diseases. The anti-oxidant, anti-carcinogenic, anti-inflammatory and apoptotic effects of these compounds have been assessed in various *in vitro* and *in vivo* systems. Since little is known about the effects of curcumin and resveratrol on FLS derived from RA patients, we investigated possible anti-inflammatory and apoptotic properties of curcumin and resveratrol in FLS obtained from two RA patients.

Our data demonstrate that curcumin and resveratrol effectively suppressed IL-1 β and PMA-induced IL-6 and VEGF-A expression in FLS derived from RA patients. Furthermore, curcumin and resveratrol induced activation of different caspases and induced apoptotic events in FLS.

Although further studies are needed, these results provide important new insights into the biological effects of curcumin and resveratrol, and suggest both substances as natural remedies for the treatment of chronic inflammatory diseases like RA.

References

1. Verhagen AP, de Vet HC, de Bie RA, Kessels AG, Boers M, Knipschild PG (2000). Balneotherapy for rheumatoid arthritis and osteoarthritis. *Cochrane Database Syst Rev* CD000518.
2. Stuhlmeier KM, Bröll J, Iliev B (2009) NF-kappaB independent activation of a series of pro-inflammatory genes by hydrogen sulfide. *Exp Biol Med* (Maywood) 234(11):1327-38.
3. Whiteman M, Li L, Rose P, Tan CH, Parkinson DB, Moore PK (2010) The effect of hydrogen sulfide donors on LPS-induced formation of inflammatory mediators in macrophages. *Antioxid Redox Signal* Feb 3.
4. Baskar R, Li L, Moore PK (2007) Hydrogen sulfide induces DNA damage and changes in apoptotic gene expression in human lung fibroblast cells. *FASEB J* 21:247-55.
5. Truong DH, Eghbal MA, Hindmarsh W, Roth SH, O'Brien PJ (2006) Molecular mechanisms of hydrogen sulfide toxicity. *Drug Metab Rev* 38:733-44.
6. Szabó C, Kiss L, Pankotai E (2007) Cytoprotective and anti-inflammatory effects of hydrogen sulfide in macrophages and in mice. *Crit Care* 11(Suppl.2),P2.

7. Kimura Y, Dargusch R, Schubert D, Kimura H (2006) Hydrogen sulfide protects HT22 neuronal cells from oxidative stress. *Antioxid Redox Signal* 8:661-70.
8. Whiteman M et al. (2004) The novel neuromodulator hydrogen sulfide: an endogenous peroxynitrite 'scavenger'? *J Neurochem* 90:765-68.
9. Whiteman M et al. (2005) Hydrogen sulfide: a novel inhibitor of hypochlorous acid mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 326:794-98.
10. Yan SK et al. (2006) Effects of hydrogen sulfide on homocysteine-induced oxidative stress in vascular smooth muscle cells. *Biochem Biophys Res Commun* 361:485-91.
11. Nagai Y, Tsugane M, Oka J, Kimura H (2004) Hydrogen sulfide induces calcium waves in astrocytes. *FASEB J* 18:557-59.
12. Yang G, Wu L, Wang R (2006) Pro-apoptotic effect of endogenous H₂S on human aorta smooth muscle cells. *FASEB J* 20:553-55.
13. Kloesch B, Liszt M, Broell J (2010) H₂S transiently blocks IL-6 expression in rheumatoid arthritic fibroblast-like synoviocytes and deactivates p44/42 mitogen-activated protein kinase. *Cell Biol Int* 34(5):477-84.
14. Kloesch B, Liszt M, Steiner G, Broell J (2012) Inhibitors of p38 and ERK1/2 MAPkinase and hydrogen sulphide block constitutive and IL-1 β -induced IL-6 and IL-8 expression in the human chondrocyte cell line C-28/I2. *Rheumatol Int* 32:729-36.
15. Kloesch B, Liszt M, Krehan D, Broell J, Kiener H, Steiner G (2012) High concentrations of hydrogen sulphide elevate the expression of a series of pro-inflammatory genes in fibroblast-like synoviocytes derived from rheumatoid and osteoarthritis patients. *Immunol Lett* 141:197-203.
16. Santos NC, Figueira-Coelho J, Martins-Silva J, Saldanha C (2003) Multidisciplinary utilization of dimethyl sulfoxide: pharmacological, cellular, and molecular aspects. *Biochem Pharmacol* 65:1035-41.
17. Pearson TW, Dawson HJ, Lackey HB (1981) Natural occurring levels of dimethyl sulfoxide in selected fruits, vegetables, grains, and beverages. *J Agric Food Chem* 29:1089-91.
18. Jacobs S, Lawrence RM, Siegel M (1999) *The Miracle MSM: the Natural Solution for Pain*. Putnam GP, New York.
19. Kim LS, Axelrod LJ, Howard P, Buratovich N, Waters RF (2006) Efficacy of methylsulfonylmethane (MSM) in osteoarthritis pain of the knee: a pilot clinical trial. *Osteoarthritis Cartilage* 14:286-94.
20. Goldring MB (2004) Culture of immortalized chondrocytes and their use as models of chondrocyte function. *Methods Mol Med* 100:37-52.
21. Kloesch B, Liszt M, Broell J, Steiner G (2011) Dimethyl sulphoxide and dimethyl sulphone are potent inhibitors of IL-6 and IL-8 expression in the human chondrocyte cell line C-28/I2. *Life Sci* 89:473-8.
22. Aggarwal BB, Kumar A, Bharti AC (2003) Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 23:363-98.
23. Park C, Moon DO, Choi IW, Choi BT, Nam TJ, Rhu CH, Kwon TK, Lee WH, Kim GY, Choi YH (2007) Curcumin induces apoptosis and inhibits prostaglandin E(2) production in synovial fibroblasts of patients with rheumatoid arthritis. *Int J Mol Med* 20:365-72.
24. Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, Fong HH, Farnsworth NR, Kinghorn AD, Mehta RG, et al. (1997) Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science (Wash DC)* 275:218-20.
25. Bhat KPL, Kosmeder JWII, Pezzuto JM (2001) Biological effects of resveratrol. *Antioxid Redox Signal* 3:1041-64.
26. Dorrie J, Gerauer H, Wachter Y, Zunino SJ (2001) Resveratrol induces extensive apoptosis by depolarizing mitochondrial membranes and activating caspase-9 in acute lymphoblastic leukemia cells. *Cancer Res* 61:4731-39.